

Lizard Room Daily Tasks

Spraying

Spray each cage with RO water, set nozzle to the “mist” setting. Try to spray onto walls of cage and avoid saturating soil.

Morning spray should be done before **10AM**

Afternoon spray after **4PM**

Incubating Eggs

Check incubating eggs for hatchlings or failed incubations (see dead animal procedure).

Record the hatch date in the data log. Open the egg cup in a butterfly cage and gently capture the hatchling. Toe clip following the charts posted in the lizard room. Set up the hatchling in the next available cage. Cages must have no more than 6 animals, which must have hatched within 7 days of each other. Label the animal's number, cross type, hatch date and originating cage on the hatchling cage.

Feeding Babies

Use a deli cup to collect pinhead crickets, from tub in lab. Leave tub outside of lizard room to slow cricket growth. Refill cricket food and water crystals if necessary. Dust pinheads with light coating of appropriate vitamin powder; see whiteboard as these vary by day.

Check on every hatchling and remove any dead hatchlings (see dead animal procedure) prior to feeding. Feed 6-10 pinhead per hatchling, be sure pinheads are small enough to be eaten by smallest animal in the cage, (e.g. small enough to fit in its mouth).

Dead Animal Procedure

It is critically important that you properly identify and label dead animals. It is very difficult to undo mistakes made at this stage.

Failed eggs

Remove egg and clean off any stuck vermiculite. Record death date in data log. Using forceps and dissection scissors dissect embryo from failed egg. Store in 1.5 mL tube with egg number labeled on the outside in sharpie and inside the tube in pencil on a piece of paper. Put egg in the next available slot in the hatchling box in the -80.

Dead Animals

Carefully identify animal using toe clipping. If there is any ambiguity in the toe clipping due to decay, identify every other animal remaining in the cage and determine the number of the dead animal by elimination. Using a single line to cross off the dead animal from the cage, leaving the information legible.

Hatchlings: Remove hatchling from cage. Record death date in data log. Store in 1.5 mL tube with hatchling number labeled on the outside in sharpie and inside the tube in pencil on a piece of paper.

Adults: Determine which experiment the animal came from. Enter death data for Julianne's animals in the Blue binder, F1_s in the red binder and Anthony's P1_s (parents of the F1_s) on sheets in the front pocket of the blue binder.

F1_s and Julianne's animals should be placed in a 15mL tube with the specimen number labeled on the outside in sharpie and inside the tube in pencil on a piece of paper. Store tube in the next available box in the -80.

P1 animals have been assigned Glor numbers; the tags for these animals are in the second right-side drawer. Tie the appropriate tag around the waist of the animal and store in a labeled 15mL tube in the next available box in the -80.